

A genetic locus and gene expression patterns associated with the priming effect on lettuce seed germination at elevated temperatures

Andrés R. Schwember · Kent J. Bradford

Received: 4 September 2009 / Accepted: 16 December 2009 / Published online: 3 January 2010
© The Author(s) 2010. This article is published with open access at Springerlink.com

Abstract Seeds of most cultivated varieties of lettuce (*Lactuca sativa* L.) fail to germinate at warm temperatures (i.e., above 25–30°C). Seed priming (controlled hydration followed by drying) alleviates this thermoinhibition by increasing the maximum germination temperature. We conducted a quantitative trait locus (QTL) analysis of seed germination responses to priming using a recombinant inbred line (RIL) population derived from a cross between *L. sativa* cv. Salinas and *L. serriola* accession UC96US23. Priming significantly increased the maximum germination temperature of the RIL population, and a single major QTL was responsible for 47% of the phenotypic variation due to priming. This QTL collocated with *Htg6.1*, a major QTL from UC96US23 associated with high temperature germination capacity. Seeds of three near-isogenic lines (NILs) carrying an *Htg6.1* introgression from UC96US23 in a Salinas genetic background exhibited synergistic increases in maximum germination temperature in response to priming. *LsNCED4*, a gene encoding a key enzyme (9-*cis*-epoxycarotenoid dioxygenase) in the abscisic acid biosynthetic pathway, maps precisely with *Htg6.1*. Expression of *LsNCED4* after imbibition for 24 h at high temperature was greater in non-primed seeds of Salinas, of a second cultivar (Titan) and of NILs containing *Htg6.1* compared to primed seeds of the same genotypes. In contrast, expression of genes encoding regulated enzymes in the gibberellin and ethylene biosynthetic pathways (*LsGA3ox1* and *LsACSI*,

respectively) was enhanced by priming and suppressed by imbibition at elevated temperatures. Developmental and temperature regulation of hormonal biosynthetic pathways is associated with seed priming effects on germination temperature sensitivity.

Keywords Abscisic acid · Ethylene · Germination · Gibberellin · Priming · QTL · Thermoinhibition

Introduction

Seed priming is the controlled hydration of seeds to allow completion of the first steps in the germination process before the seeds are subsequently dried, distributed and sown (Bruggink 2004). To achieve this, seeds are partially hydrated to a water content where germination processes can begin, but radicle emergence does not occur (Heydecker and Coolbear 1977). Seed water content can be controlled by priming seeds in osmotic solutions (osmopriming) or by supplying only predetermined amounts of water to the seeds or limiting the imbibition time (hydropriming). Following the hydration period, seeds are subsequently dried to their original moisture content and stored or planted. In general, seed priming induces faster and more uniform germination over a broader temperature range when the primed seeds are subsequently planted (McDonald 2000). This practice is applied commercially in many crops to improve germination rate (speed), and particularly in lettuce (*Lactuca sativa* L.) to alleviate thermoinhibition (failure of seeds to germinate when imbibed at warm temperatures) by increasing the maximum temperature at which germination will occur (Cantliffe et al. 1981; Valdes et al. 1985).

Little is known about the genetics of seed priming or the potential genes or mechanisms involved in increasing the

Electronic supplementary material The online version of this article (doi:10.1007/s11103-009-9591-x) contains supplementary material, which is available to authorized users.

A. R. Schwember · K. J. Bradford (✉)
Department of Plant Sciences, One Shields Ave., University
of California, Davis, CA 95616-8780, USA
e-mail: kjbradford@ucdavis.edu

maximum germination temperature of primed seeds. Soeda et al. (2005) found that a small but distinct group of seed maturation genes (i.e., late-embryogenesis abundant and storage compound-related genes) correlated to stress tolerance in primed seeds of *Brassica oleracea*. Those authors also discovered that a large group of genes was up-regulated during germination in water, but not during priming, representing genes that are specific to germination in water or are inhibited by low water potential. Gao et al. (1999) found that transcripts of two different aquaporins (*BnPIP1* and *Bn γ -TIP2*) were present earlier during germination of primed canola (*Brassica napus*) seeds than of non-primed seeds, although priming itself induced only the expression of *BnPIP1*. They proposed that *BnPIP1* is related to the water transportation required for enzymatic metabolism of storage nutrients at the early stages of canola seed germination while *Bn γ -TIP2* expression is related to cell growth associated with radicle protrusion. In *Arabidopsis*, Gallardo et al. (2001) identified three priming-associated proteins (i.e., one tubulin subunit and two 12S-cruciferin β -subunits) whose abundance or solubility increased during priming treatments. An increase in 11-S globulins associated with storage protein mobilization has been reported in primed sugar beet (*Beta vulgaris*) seeds (Job et al. 1997), whereas tubulin subunits are presumably involved in reactivation of cell cycle activity during priming (de Castro et al. 2000). In addition, two low-molecular weight heat shock proteins specifically increased during osmopriming (Gallardo et al. 2001). Ligterink et al. (2007) identified a number of genes in tomato (*Solanum lycopersicum*) whose expression is associated either positively or negatively with responses to priming, but their modes of action have not been determined. Nascimento et al. (2001) showed that endo- β -mannanase activity increased in the lettuce endosperm during priming and before or coincident with radicle emergence in non-primed seeds, and suggested that it may be involved in weakening of the micropylar endosperm to allow radicle emergence. In addition, seeds of a thermotolerant lettuce variety (Everglades) exhibited much higher endo- β -mannanase activity than did seeds of a thermosensitive variety (Dark Green Boston). Similarly, Dutta et al. (1997) reported that activity of a cell wall-bound endo- β -mannanase extracted from lettuce endosperms prior to radicle emergence was reduced by imbibition at high temperature. However, others have reported that endo- β -mannanase is present only in the lateral endosperm after radicle emergence in lettuce seeds (Halmer et al. 1975; Nonogaki and Morohashi 1999). Thus, the specific genetic or molecular mechanisms underlying the increase in the maximum germination temperature by seed priming remain unclear.

Abscisic acid (ABA), gibberellins (GA), and ethylene have been implicated in regulating germination and

dormancy in lettuce seeds in relation to light and temperature, with ABA inhibiting and GA and ethylene promoting germination (Cantliffe et al. 2000; Kucera et al. 2005). A key regulated step in ABA biosynthesis is catalyzed by 9-*cis*-epoxycarotenoid dioxygenase (NCED), which cleaves 9-*cis*-xanthophyll to xanthoxin, a precursor of ABA (Tan et al. 2003; Lefebvre et al. 2006). Our group previously identified a quantitative trait locus (QTL) for high temperature germination (*Htg6.1*) conferred by *L. serriola* accession UC96US23, one parent of a recombinant inbred line (RIL) population derived from a cross with *L. sativa* cv. Salinas (Argyris et al. 2005, 2008b). One member of the NCED gene family in lettuce (*LsNCED4*) is regulated by photochrome, being induced by far-red light and repressed by red light (Sawada et al. 2008a). *LsNCED4* maps to the same genetic region as *Htg6.1* and is induced by imbibition at high temperature in genotypes susceptible to thermoinhibition, resulting in elevated seed ABA content (Argyris et al. 2008a). The gene encoding an enzyme that catalyzes the last step in the formation of active GA₁, GA 3- β -hydroxylase (*LsGA3ox1*, formerly called *Ls3h1*) is a control point in the regulation of GA biosynthesis by light in lettuce seeds (Toyomasu et al. 1998; Sawada et al. 2008b). Expression of *LsGA3ox1* increases during imbibition in the light prior to radicle emergence, but is repressed in lettuce seeds imbibed at high temperature (Argyris et al. 2008a), possibly through the action of ABA (Sawada et al. 2008b; Toh et al. 2008). Conversely, both GA-dependent and GA-independent mechanisms can influence ABA metabolism in lettuce seeds (Sawada et al. 2008a). Ethylene production by lettuce seeds is correlated positively with the ability to germinate at high temperature (Saini et al. 1986, 1989; Nascimento et al. 2000; Kozarewa et al. 2006). Expression of the 1-aminocyclopropane carboxylate (ACC) synthase gene (*LsACSI*), which encodes the enzyme catalyzing the rate-limiting step of ethylene biosynthesis, is suppressed in lettuce seeds by imbibition at high temperatures (Argyris et al. 2008a). Thus, imbibition of lettuce seeds at high temperature results in promotion of ABA biosynthesis and repression of GA and ethylene biosynthesis, leading to inhibition of germination. However, it is not known whether seed priming alleviates thermoinhibition by reducing the expression of *LsNCED4*, resulting in decreased synthesis of ABA, or by promoting the expression of *LsGA3ox1* or *LsACSI*, increasing GA and ethylene synthesis, either of which may enhance the germination of lettuce seeds at high temperatures.

The objectives of this study were to identify genetic loci associated with the alleviation of thermoinhibition of lettuce seeds due to priming and to determine whether the expression of genes associated with ABA, GA and ethylene biosynthesis during seed imbibition were altered by priming. We conducted a QTL analysis of the ability of priming

to increase the maximum temperature of lettuce seed germination. In addition, near-isogenic lines (NILs) derived by introgressing a genomic region containing a QTL associated with the capacity to germinate at high temperature (*Htg6.1*) into the Salinas genetic background were analyzed for their responses to priming. Expression of *LsNCED4*, *LsGA3ox1* and *LsACSI* was assessed in relation to the effect of priming on the maximum temperature for germination. The results support key roles for suppression of *LsNCED4* expression and for enhancement of *LsGA3ox1* and *LsACSI* expression in the alleviation of thermoinhibition by seed priming.

Materials and methods

Genotypes and seeds

A recombinant inbred line (RIL) population derived from an interspecific cross between *L. sativa* cv. Salinas and *L. serriola* accession UC96US23 that has been analyzed previously for seed germination and quality traits was utilized (Argyris et al. 2005, 2008b). This lettuce mapping population was developed by Dr. Richard Michelmore's group at UC Davis (Truco et al. 2007). Near-isogenic lines (NILs) were developed by backcrossing a RIL carrying the UC96US23 allele at *Htg6.1* into the recurrent parent Salinas (Argyris 2008). Three generations of backcrossing with marker-assisted selection followed by two selfing generations allowed the identification of NILs homozygous for an introgression containing *Htg6.1* from UC96US23 but >95% Salinas in the remaining genome (Argyris 2008). The size of the introgression varied among NIL lines, but represented roughly half of linkage group 6. In Figs. 4 and 6, the sub-NILs pooled for each genotype were the following: NIL 2 lines homozygous for the Salinas (133, 144, 165, 179, and 190) or UC96US23 (109, 114, 163, 193, and 198) allele; NIL 27 lines homozygous for the Salinas (105, 163, 172, 180, and 249) or UC96US23 (112, 117, 128, 131, and 143) allele; and NIL 86 lines homozygous for the Salinas (183, 187, 188, 192, and 206) or UC96US23 allele (132, 133, 151, 159, and 169) (Argyris 2008). Seeds of both the RIL populations and the NIL families were produced in the field in Davis, California in 2006. A commercial lettuce cultivar (Titan) was a selection from and therefore has a very similar genetic background as Salinas; seeds were provided by Synergene Seed & Technology, Inc., Salinas, CA.

Seed priming protocol

Seeds of the parent lines, 89 RILs, and 3 NILs were osmoprimed using a standard protocol (Heydecker et al.

1973; Tarquis and Bradford 1992) as follows: 1.2 g of seeds of each line was placed in an 8.8-cm Petri dish and the seeds were primed over one blotter in 10 mL of water or -1.25 MPa solution of polyethylene glycol (PEG) 8000 for 48 h at 9°C under continuous fluorescent light. Seeds were then rinsed briefly with water and surface water was removed by suction in a Buchner funnel. Subsequently, seeds were rapidly dried for 2 h at 32°C and 25–30% relative humidity (RH) in a ventilated oven, then were transferred to a chamber at 9°C and 30% RH and equilibrated for at least 2 days to constant moisture content. Seeds were stored thereafter in the same condition. Titan seeds (50 g) were osmoprimed in 250 mL of aerated -1.25 MPa solution of PEG 8000 at 9°C under continuous fluorescent light for durations of 6, 12, 24 and 48 h. Subsequently, the same post-priming drying and storage conditions were employed as described above.

Germination assays

Tests for germination were conducted on filter papers moistened with 4 mL of H₂O in 3.8-cm diameter Petri dishes in constant temperature chambers or on a temperature gradient table with continuous fluorescent light. Total germination (radicle emergence) was scored 3 days after planting.

A thermogradient table was used to test germination at 1°C increments across the following temperature ranges: for Salinas seeds, from 25 to 35°C; for UC96US23 seeds, from 34 to 44°C; and for the RIL and NIL populations, from 27 to 35°C. In all these tests, two replicates of 25 seeds each were placed on two blotters wetted with 4 mL of distilled water in 3.8-cm Petri dishes and covered with tight-fitting lids. The percentage of radicle emergence was assessed 48 h after planting for the RILs and the NILs, and 72 h after planting for Salinas and UC96US23.

The germination percentages of control and primed seeds of the RIL population and of the sub-lines of NILs 2, 27 and 86 were transformed to probits to normalize variances in data distribution. Statistical analysis of the overall priming effect was conducted on the probit-transformed values using the General Linear Model of the SAS statistical program (SAS Institute Inc., Cary, NC, USA).

QTL analyses

For low resolution mapping, a set of 486 markers approximately 2–3 cM apart on nine chromosomal linkage groups were chosen as the framework map for QTL analysis (Argyris et al. 2005). For QTL associated with increased germination at high temperature following priming, the differences between probit-transformed germination percentage values for control and primed seeds of

each RIL were analyzed using composite interval mapping (CIM) in Windows QTL Cartographer V. 2.5. QTL were verified by 1,000 permutations of phenotypic data and declared significant ($P = 0.05$) above the permuted log likelihood ratio (LOD score) threshold of 3.2.

For high-resolution mapping, markers from a high density Affymetrix GeneChip® mapping array (van Leeuwen et al. 2007) were combined with the framework markers comprising linkage group 6 from the *Lactuca* integrated map (Truco et al. 2007). Sequence polymorphisms revealed by differential hybridization of DNA from 113 RILs hybridized in duplicate to the array were ordered using MadMapper (<https://cgpdb.ucdavis.edu/XLinkage/MadMapper>) (van Leeuwen et al. 2009). A marker was selected from each distinct genetic bin (unique recombinations between adjacent markers) covering linkage group 6 and these were entered into QTL Cartographer to identify relevant sets of genetic markers encompassing the QTL interval. These, plus the framework markers from the integrated map, were entered into JoinMap V 4.0 (Stam 1995) and the resulting maps with genetic distances were then reanalyzed in QTL Cartographer. A covariance analysis was performed to adjust the germination values for the effects of the *Htg6.1* alleles (either Salinas or UC96US23). The differences between probit-transformed germination percentage values for control and primed seeds of each RIL were corrected by the allelic effects using a covariate regression analysis (using PROC REG and PROC GLM of SAS), and adjusted values were subsequently analyzed by QTL Cartographer and ANOVA.

Gene expression analyses

Total RNA was extracted from 0.5 g of whole Salinas seeds at 0, 8, and 24 h after imbibition at 32°C (Fig. 5), and from 0.5 g of whole Titan seeds primed for 0, 6, 12, 24, and 48 h followed by drying and after subsequent imbibition at 35°C (Fig. 7) using methods previously described (Cooley et al. 1999; Argyris et al. 2008a). In Fig. 6, multiple lines were pooled to randomize the genetic background at locations other than the introgression containing *Htg6.1*. Twenty seeds of each of 5 BC₃S₂ sub-NILs described above (100 seeds in total, which constituted one biological replicate) of NILs 27 and 86 were pooled by genotype at the *Htg6.1* marker locus: Salinas alleles were pooled separately from UC96US23 alleles. Quantitative RT-PCR assays of gene expression were conducted using methods and normalization procedures described previously (Argyris et al. 2008a). The genes assayed in this study are *LsNCED4*, *LsGA3ox1* and *LsACS1* along with *18s1*, *PP2A*, and *UBC21* for normalization; the accession and contig numbers and primer combinations used are shown in Supplemental Table SI of Argyris et al. (2008a).

Results

Priming increases the maximum germination temperature of lettuce seed

Priming seeds of Salinas and UC96US23 increased their germination at warm temperatures relative to the respective control (non-primed) seeds (Fig. 1). Before priming, over 95% of Salinas seeds germinated up to 27°C, but only 2% germinated at 29°C. In contrast, the primed seeds exhibited >90% germination up to 30°C, with a more gradual decline with increasing temperatures, reaching 0% at 35°C. The median maximum temperature for germination (T_{50} , or the temperature at which germination is reduced to 50%) increased by 4°C due to priming. The maximum germination temperature of UC96US23 seeds also increased due to priming, but since the control seeds already germinated fully to 35°C and were not completely inhibited until 38°C, the potential for increasing the maximum was limited (Fig. 1). Nonetheless, germination of the primed seeds remained greater than 90% as high as 37°C, an increase of 1–2°C compared to the control UC96US23 seeds.

Priming effects on maximum germination temperature associate with a QTL

Seeds of 89 recombinant inbred lines (RILs) derived from a cross between Salinas and UC96US23 were left untreated (control) or primed and imbibed at temperatures between 27 and 35°C to determine their maximum germination temperatures. Priming increased the maximum germination temperature of about half of the RILs (Fig. 2). Statistical analyses conducted using ANOVA confirmed that priming

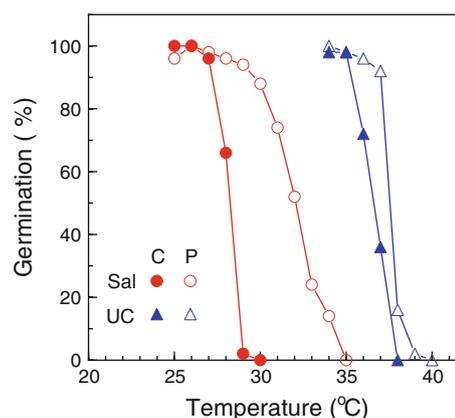
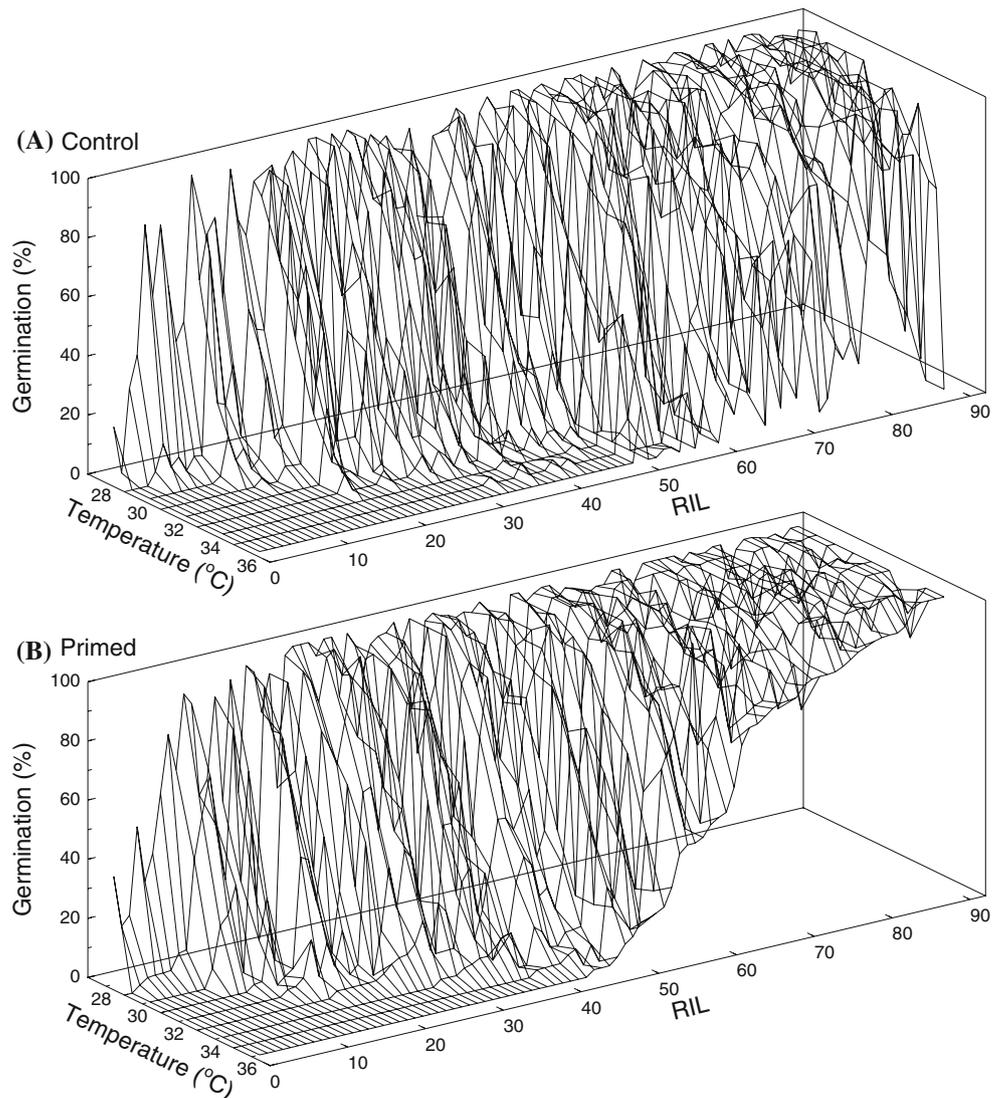


Fig. 1 Sensitivity of seed germination of *Lactuca sativa* cv. Salinas (Sal, circles) and *L. serriola* UC96US23 (UC, triangles) to temperature and priming (untreated, closed symbols; primed, open symbols). Seeds (25 in each of two replicates) were imbibed in Petri dishes at each temperature and radicle emergence was scored 72 h after planting

Fig. 2 Temperature responses of germination of control (a) and primed (b) seeds of 89 recombinant inbred lines (RILs) at 27–35°C. Two replicates of 25 seeds each were tested at each temperature on a gradient table, and radicle emergence was scored 48 h after planting. In both panels, RILs are ranked in order of increasing germination percentage of the primed seeds at the maximum temperature at which germination occurred

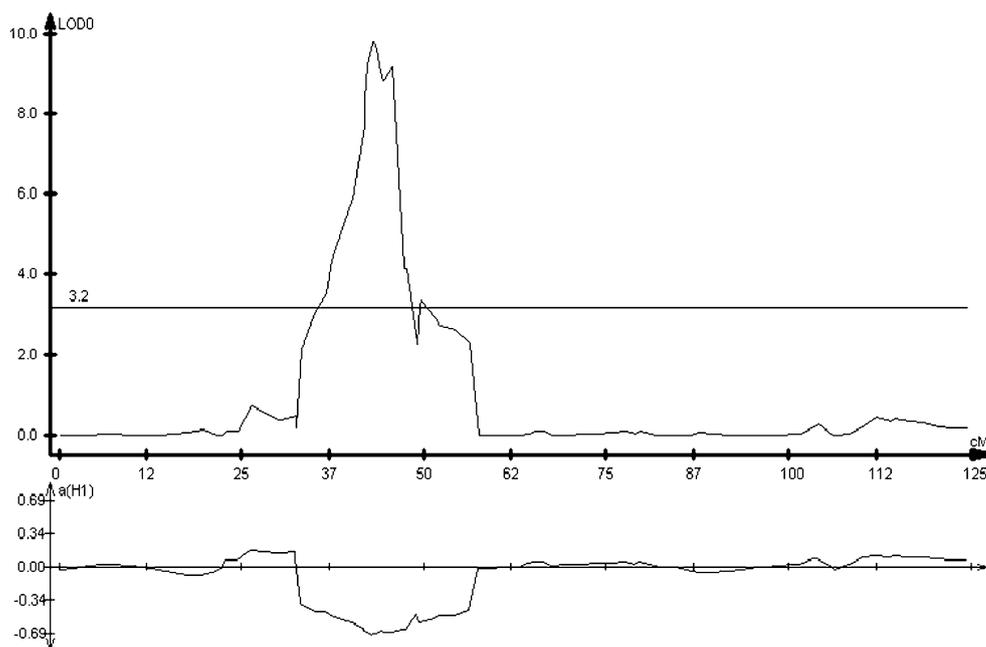


significantly increased germination of the RIL population ($P < 0.0001$) (Supplemental data, Table S1). For example, primed seeds overall exhibited 61.9% germination versus 49.7% for control seeds at 30°C and 31.5% versus 13.1% at 35°C. In addition, a significant interaction among RILs, priming, and temperature was detected ($P < 0.0001$) (Table S1), confirming that RILs responded differently to priming and to temperature across the 27–35°C temperature gradient. Seeds of RILs that germinated better at warm temperatures also showed the largest increases in germination due to priming (Fig. 2).

To investigate the genetic basis of the germination increase by seed priming, QTL analyses were performed on the probit-transformed germination percentage data for control and primed seeds and for the increase in germination due to priming (probit of primed percentage minus probit of control percentage). Using low resolution mapping, a single significant QTL was identified on linkage

group (chromosome) 6 that was responsible for 39.3% of the variance and was conferred by the UC96US23 allele at this locus (data not shown). This QTL collocated with one responsible for over 60% of the phenotypic variation in high temperature germination between the parent lines, provided also by the UC96US23 allele, termed *Htg6.1* (Argyris et al. 2005, 2008b). High resolution mapping on linkage group 6 was subsequently conducted utilizing a high-density map based on sequence-based polymorphisms detected using a tiled microarray representing over 35,000 lettuce genes (van Leeuwen et al. 2009). Again, a single QTL was identified that was responsible for 46.9% of the variation in the increase in germination at 35°C due to priming (Fig. 3). This QTL again collocated precisely with the QTL for high temperature germination *Htg6.1*, and *LsNCED4* mapped to the center of the QTL confidence interval (Fig. 3). Similar results were found using only the control or only the primed germination data (not shown).

Fig. 3 High-resolution mapping of a QTL on chromosome 6 associated with the increase in maximum germination temperature due to priming when tested at 35°C using a RIL population derived from *L. sativa* cv. Salinas and *L. serriola* UC96US23. The LOD (log of odds) scores in the upper graph are significant above a value of 3.2 (horizontal line). The lower graph indicates which parent contributed the trait, with positive values indicating Salinas and negative values indicating UC96US23. The QTL confidence interval spans the chromosomal region from 33 to 56 cM. The maximum QTL peak exhibits a 9.8 LOD score positioned at 43 cM, collocating with *LsNCED4* at 43.5 cM



Further fine-mapping has reduced the *Htg6.1* confidence interval to as little as 1 cM in some seed production environments, and *LsNCED4* remains centered in the *Htg6.1* interval (Argyris et al., unpublished results). Thus, the *Htg6.1* QTL that confers germination thermotolerance in this population is also associated with the increase in high temperature germination following priming.

As *Htg6.1* had a large effect on maximum germination temperature in both primed and control seeds, it may be masking loci having smaller effects in the statistical analysis. We therefore conducted a covariate analysis to remove the effect of *Htg6.1* and determine whether other QTL were present. Even after controlling for the effect of the *Htg6.1* allele, the RIL \times Treatment \times Temperature interaction was highly significant, indicating that RILs still responded differently to priming across temperatures (Table S2). When germination values adjusted for the effect of the *Htg6.1* allele across the entire population were analyzed only for control seeds, no QTL was detected, confirming that the covariate analysis had corrected for the effect of *Htg6.1*. However, QTL mapping using the primed seed data or the difference in germination between control and primed seeds identified the *Htg6.1* locus as highly significant (LOD score 8.2, 26.1% of remaining variation explained) as well as a second minor QTL on chromosome 5 that just exceeded the LOD score threshold (LOD score 3.5, 11.3% of the remaining variation explained). However, the second QTL was not confirmed by high-resolution mapping (LOD = 2.0; data not shown). Thus, the UC96US23 *Htg6.1* locus conditions a significant priming response that increases germination at high temperature beyond its effect in control seeds. We were unable to

conclusively identify other QTL that might contribute to this effect.

Responses to priming of near-isogenic lines containing Salinas or UC96US23 alleles at *Htg6.1*

Marker-assisted backcrossing and selection were utilized to introgress a genomic region containing *Htg6.1* from UC96US23 into a Salinas background and to develop NILs (Argyris et al. 2008b; Argyris 2008). Five BC₃S₂ NIL progeny lines derived from each of three NIL families (2, 27 and 86) were assessed to test whether this genomic region is specifically associated with differential responses to priming. Control or primed seeds from progeny NILs homozygous for either the Salinas or the UC96US23 alleles at *Htg6.1* were tested for germination at high temperatures (Fig. 4). Without priming, the presence of the UC96US23 alleles increased the T_{50} by about 2°C, consistent with other results for these NILs (Argyris 2008). Surprisingly, NILs with Salinas alleles at *Htg6.1* showed a reduced response to priming compared to that of the Salinas parent (compare Figs. 1, 4). The increase in the median temperature for germination by priming in NILs 27 and 86 was negligible, and less than 1°C for NIL 2 (Fig. 4). As responses to priming can vary among different seed lots of the same variety dependent upon production environment, storage conditions, and age, responses to priming of the NILs homozygous for the Salinas allele at *Htg6.1* might have been enhanced by testing variations in the priming protocol. However, using the same priming conditions, progeny seeds of the same NILs homozygous for the UC96US23 allele exhibited strong responses to priming,

with increases in their T_{50} values of 2.6, 3.1 and 3.5°C for NILs 2, 27, and 86, respectively, compared to the corresponding control seeds (Fig. 4). Primed seeds of NILs containing the UC96US23 allele at *Htg6.1* germinated at least 50% at 33°C, while seeds of the other genotypes and treatments germinated less than 10% at that temperature. Statistical analyses conducted using ANOVA confirmed that seeds of all three NILs responded differently to priming depending upon the *Htg6.1* allele present ($P < 0.0001$), and that the germination response to the

treatments (primed or non-primed) across the temperature gradient also depended upon the *Htg6.1* allele present ($P < 0.0001$) (Table S3). Although the introgressed region was fairly large due to a limited number of markers available for selection in this region at the time, these similar results in multiple NIL families (in which other remaining UC96US23 loci would be randomized) confirm that the introgressed region containing *Htg6.1* strongly influences response to priming.

Priming alters the expression of *LsNCED4*, *LsGA3ox1*, and *LsACSI* in seeds imbibed at high temperature

LsNCED4 collocates with *Htg6.1*, and higher T_{50} values were associated with reduced expression of this gene in seeds imbibed at high temperature (Argyris et al. 2008a). To determine whether priming reduces the expression of *LsNCED4* in seeds imbibed at high temperature, mRNA abundance of *LsNCED4* was analyzed in the priming-responsive Salinas parent line. Dry control seeds had 3.4-fold greater abundance of *LsNCED4* mRNA than the dry primed seeds, indicating that *LsNCED4* mRNA abundance was reduced during the priming treatment (Fig. 5a), as it is during early imbibition of control seeds (Argyris et al. 2008a). The abundance of *LsNCED4* mRNA decreased further in both control and primed seeds after 8 h of imbibition at 32°C, then increased significantly (sixfold) only in the control seeds after 24 h of imbibition. Conversely, dry primed seeds had considerably greater *LsGA3ox1* and *LsACSI* mRNA abundance than did the dry control seeds (Fig. 5b, c), indicating that those transcripts accumulated during the priming treatment. After 24 h of imbibition at 32°C, abundance of *LsGA3ox1* and *LsACSI* mRNAs remained very low or undetectable in the control seeds, but had increased fourfold to fivefold in the primed seeds.

Influence of *Htg6.1* allele on expression of *LsNCED4*, *LsGA3ox1*, and *LsACSI* in response to priming and imbibition temperature

To study how the parental *Htg6.1* alleles affected differential gene expression between primed and non-primed seeds, experiments were conducted using seeds of pooled progeny of NILs 27 and 86 homozygous for either the Salinas or UC96US23 alleles at *Htg6.1*. Preliminary tests determined the T_{50} temperatures for control and primed seeds of each seed pool and germination percentages at 25 and 35°C (Table 1). Germination was greater than 90% in all seed lots at 25°C, whereas only primed seeds germinated at 35°C, particularly those with the UC96US23 allele. The increase in T_{50} by priming was also greater in NILs carrying the UC96US23 allele at *Htg6.1*. After 24 h

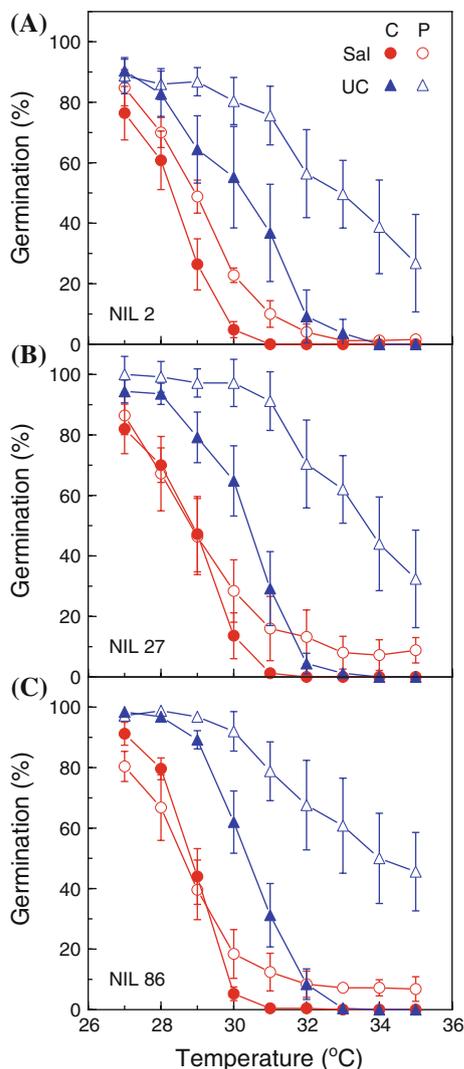


Fig. 4 Sensitivity of germination of control (*closed symbols*) and primed (*open symbols*) seeds of near-isogenic lines (NILs) 2 (*a*), 27 (*b*), and 86 (*c*) in a Salinas background introgressed with a genomic region containing *Htg6.1* QTL alleles from Salinas (Sal, *circles*) or from UC96US23 (UC, *triangles*). Five BC₃S₂ sub-NILs derived from NILs 2, 27, and 86 homozygous for either Salinas or UC96US23 alleles were assessed for germination at 27–35°C. Seeds (two replicates of 25 each) were imbibed at each temperature and radicle emergence was scored 48 h after planting. Pooled standard errors (SE) from the five sub-NILs are plotted as error bars at each temperature

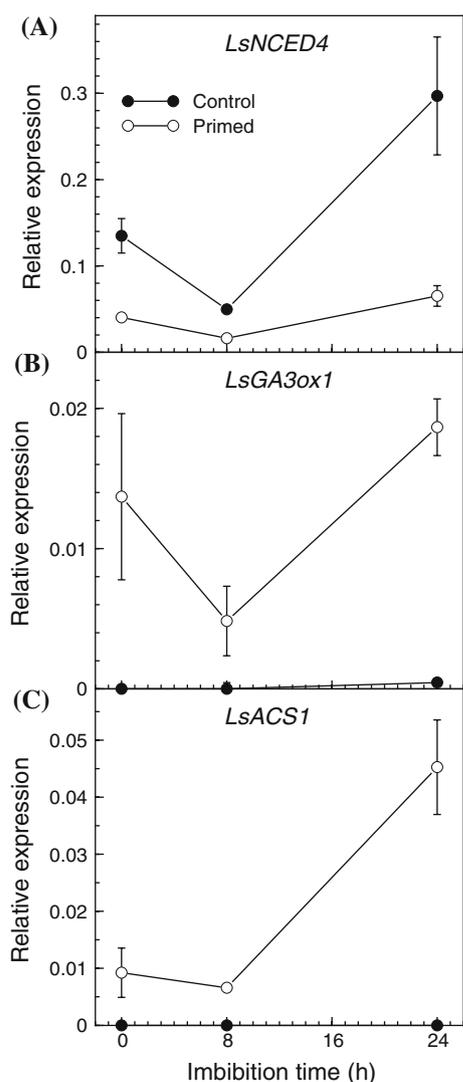


Fig. 5 Relative mRNA abundance (measured by quantitative RT-PCR) of *LsNCED4* (a), *LsGA3ox1* (b), and *LsACS1* (c) in control (closed symbols) or primed (open symbols) Salinas seeds that were dry (0 h) or imbibed at 32°C for 8 or 24 h. Only the primed seeds germinated at this temperature. Error bars are \pm SE of 3 biological replicates

of imbibition at 25 or 35°C, *LsNCED4* mRNA abundance was analyzed by quantitative RT-PCR. Regardless of the *Htg6.1* allele, priming reduced expression of *LsNCED4* in seeds imbibed at both temperatures, but especially at 35°C (Fig. 6a, b). Seeds of both NILs responded similarly, with the increase in *LsNCED4* expression in control seeds at 35°C tending to be greater in seeds carrying the Salinas allele compared to those with the UC96US23 allele.

LsGA3ox1 and *LsACS1* mRNA abundances were also assayed in the same samples. In general, expression of *LsGA3ox1* was reduced by imbibition at 35°C compared to 25°C, with very low mRNA abundance in seeds with the Salinas allele at *Htg6.1* imbibed at 35°C (Fig. 6c, d). Priming had relatively small and inconsistent effects on

Table 1 Temperature response of germination of osmoprimed and control lettuce seeds of near-isogenic lines introgressed with Salinas or UC96US23 alleles at *Htg6.1*

NIL	Treatment	Allele	T_{50} (°C)	Germination (%)	
				25°C	35°C
27	Control	Sal	28.5	92.0	0
		UC	30.5	94.4	0
	Primed	Sal	29.0	96.4	8.8
		UC	33.5	100	32.4
86	Control	Sal	28.5	96.2	0
		UC	30.0	98.4	0
	Primed	Sal	28.5	90.4	6.8
		UC	34.0	97.2	45.6

Osmoprimed and control seeds of NILs 27 and 86 homozygous for either Salinas (Sal) or UC96US23 (UC) alleles were imbibed at temperatures between 25 and 35°C on a gradient table. Two replicates of 25 seeds each were tested at each temperature and radicle emergence was scored 48 h after planting. Median maximum germination temperatures (T_{50}) and mean germination percentages at 25 and 35°C are shown

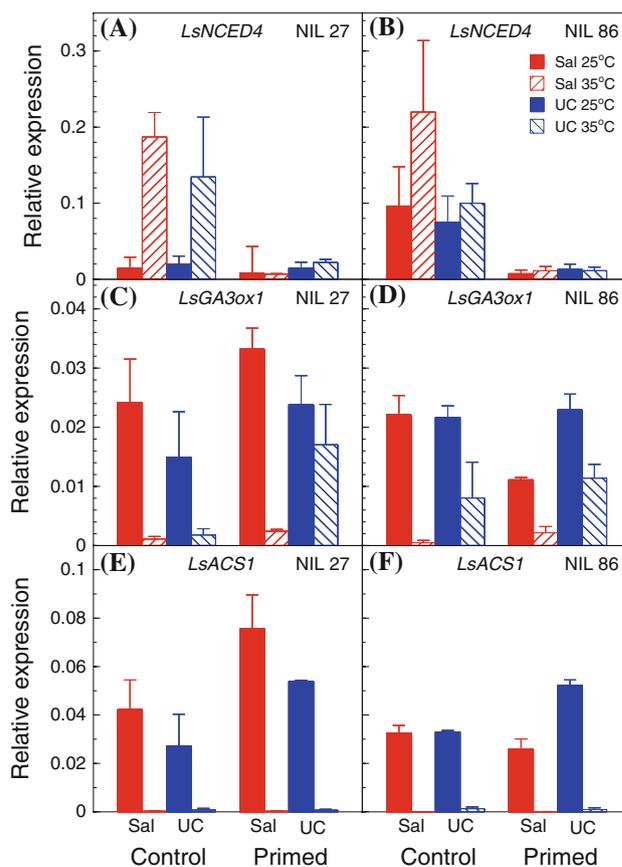


Fig. 6 Expression of *LsNCED4* (a, b), *LsGA3ox1* (c, d) and *LsACS1* (e, f) in control and primed seeds of NILs 27 (a, c, e) and 86 (b, c, f) introgressed with Salinas (Sal) or UC96US23 (UC) *Htg6.1* alleles. Seeds were imbibed at 25 (solid bars) or at 35°C (striped bars) for 24 h, then extracted. Twenty seeds of each of five BC_3S_2 sub-NILs were pooled for each *Htg6.1* allele and treatment replicate within each NIL lineage. Error bars represent \pm SE of 3 biological replicates

expression of *LsGA3ox1* between the NILs. *LsACSI* expression exhibited overall trends similar to that for *LsGA3ox1* at 25°C, but *LsACSI* expression was almost completely suppressed at 35°C in both genotypes regardless of priming treatment (Fig. 6e, f).

Expression of *LsNCED4*, *LsGA3ox1*, and *LsACSI* during and after priming, drying, and imbibition at high temperature

The effects of priming and drying on the expression of *LsNCED4*, *LsGA3ox1*, and *LsACSI* were studied using seeds of the commercial cultivar Titan, which is closely related genetically to Salinas. Germination of Titan seeds at 35°C increased with longer priming durations (Table 2), which corresponded well with declining *LsNCED4* mRNA abundance in the primed and dried seeds (Fig. 7a). *LsNCED4* mRNA abundance in the dry control seeds was 8.8-fold greater than in the dried 48-h osmoprimed seeds. In contrast, the abundance of *LsGA3ox1* and *LsACSI* mRNAs increased with longer priming durations. The dried 48-h osmoprimed seeds had 5.6-fold higher *LsGA3ox1* mRNA abundance and eightfold greater *LsACSI* mRNA abundance than the dry control seeds (Fig. 7b, c). Similar trends were observed for mRNA abundance of these genes in seeds extracted immediately after priming before drying and in hydroprimed seeds (data not shown), indicating that mRNA abundances were not affected significantly by the rapid drying following priming or by the type of priming, respectively.

The consequences of different priming durations on gene expression following imbibition at 35°C for 24 h were also assessed. *LsNCED4* mRNA abundance in control seeds and 6 or 12-h primed seeds decreased relative to the levels present before imbibition, but remained higher than in seeds primed for 24 or 48 h that germinated to higher percentages at 35°C (Fig. 7a; Table 2). Conversely, the abundance of *LsGA3ox1* and *LsACSI* mRNAs increased as

the priming times increased, particularly in the seeds primed for 24 and 48 h before imbibition at 35°C (Fig. 7b, c).

Discussion

The regulatory mechanisms controlling germination in lettuce seeds encompass networks that sense external (i.e., light and temperature) and internal (i.e., genetic and physiological) conditions and determine whether germination proceeds to completion or is inhibited prior to radicle emergence. In particular, temperature is a primary factor regulating seed germination in this species (Gonai et al. 2004; Sung et al. 2008). Lettuce seeds exhibit thermoinhibition (fail to germinate) at temperatures well below the biological upper limit for seedling growth (i.e., >28°C) (Berrie 1966; Gray 1975), and the ability of the seeds to germinate at high temperatures depends on both genetic and environmental factors (Sung et al. 1998; Kozarewa et al. 2006; Argyris et al. 2008a; Contreras et al. 2009). Thermoinhibition in lettuce can delay or prevent germination, resulting in reduced field emergence and stand establishment (Valdes et al. 1985), diminished yield (Cantliffe et al. 1981), and multiple harvests that can reduce quality and profitability (Benjamin 1990). Several plant hormones play key roles in the regulation of thermoinhibition in lettuce seeds (Saini et al. 1986; Cantliffe et al. 2000; Kucera et al. 2005). For example, exposure to ABA can decrease the maximum germination temperature, whereas exposure to GA and ethylene can increase it (Reynolds and Thompson 1973; Saini et al. 1989; Dutta and Bradford 1994; Gonai et al. 2004; Argyris et al. 2008a). Seed priming has been a valuable tool in circumventing thermoinhibition in lettuce seeds due to its ability to increase germination at high temperatures (Heydecker and Coolbear 1977; Cantliffe et al. 1981, 1984; Valdes et al. 1985; Weges 1987).

Table 2 Effect of priming in water (hydropriming) or in polyethylene glycol (PEG) 8000 (osmopriming) for different durations on germination of Titan lettuce seeds at 20 and 35°C

Type of priming	Imbibition temperature (°C)	Germination (%)				
		Priming duration at 9°C (h)				
		0	6	12	24	48
H ₂ O	20	98 ± 1	96 ± 2	97 ± 1	96 ± 2	100
	35	0	0	12 ± 2	68 ± 6	75 ± 12
PEG	20	97 ± 1	93 ± 1	100	100	96 ± 2
	35	0	0	1 ± 1	40 ± 4	88 ± 6

After priming for the indicated durations at 9°C, seeds were rapidly dried (2 h at 32°C and 30% RH) and equilibrated at 9°C and 30% RH for 2 days. Subsequently, the seeds were imbibed at 20 and 35°C and radicle emergence was scored after 48 h. Three replicates of 25 seeds were tested for each treatment; means and standard errors are indicated

The endosperm envelope surrounding the embryo constitutes a physical barrier to radicle emergence and is required for the imposition of thermoinhibition in lettuce seeds (Borthwick and Robbins 1928; Speer 1974). The rate and extent of weakening of the micropylar endosperm enclosing the radicle tip may be involved in determining whether thermoinhibition occurs (Sung et al. 1998). Sung et al. (2008) recently reported that seed priming reduced the physical resistance of the lettuce endosperm during imbibition by weakening the cell walls and by depleting stored reserves leading to cell collapse prior to radicle emergence. Alternatively, Bradford and Somasco (1994) concluded from experiments comparing intact seeds and excised embryos that the effects of priming in lettuce occurred primarily in the embryo, rather than in the surrounding endosperm tissues, consistent with the proposal by Cantliffe et al. (1984) that priming acted by initiating irreversible embryo expansion. As embryo growth is required to complete germination even if there are structural changes in the endosperm, inhibition of embryo expansion and of endosperm weakening may both be involved in thermoinhibition (Nonogaki et al. 2007). While ABA has long been known to reduce embryo growth capacity (Takeba and Matsubara 1979; Schopfer and Plachy 1984), its effects on endosperm weakening are more variable, with reports of inhibition of weakening in some cases (Toorop et al. 2000; Muller et al. 2006) and others in which relatively little effect was found (Chen and Bradford 2000; Wu et al. 2001). GA, on the other hand, generally promotes both embryo growth and endosperm weakening (Nonogaki et al. 2007).

In order to provide further insight into the causes of lettuce seed thermoinhibition, we investigated the genetic regulation of the effect of seed priming on the temperature sensitivity of germination. We identified a major QTL on chromosome 6 derived from *L. serriola* accession UC96US23 that accounted for 47% of the variance for the increase in germination at high temperature due to seed priming (Fig. 3). This QTL collocated with a previously identified QTL for high temperature germination ability (*Htg6.1*) in non-primed seeds of the same RIL population (Argyris et al. 2005, 2008b). Covariate analysis showed a significant QTL associated with priming at this locus even when the effect of the *Htg6.1* alleles on basal thermotolerance was removed. This was confirmed with seeds of NILs with Salinas as a recurrent parent that were introgressed with a genomic region containing the UC96US23 allele at this locus, which exhibited both greater high temperature tolerance and an enhanced response to priming (Fig. 4). Thus, the UC96US23 allele at *Htg6.1* increases the maximum germination temperature and the response of seeds to priming, suggesting that priming influences the same regulatory mechanisms that control the temperature sensitivity of seed germination.

Both the QTL interval and the introgressed region span hundreds of genes, but considerable evidence points to *LsNCED4* as the primary candidate gene. Additional mapping has reduced the *Htg6.1* QTL confidence interval to as little as 1 cM, and *LsNCED4* remains centered in the QTL interval (Argyris et al., unpublished results). Just as reduced expression of the UC96US23 allele of *LsNCED4* was associated with high temperature germination capacity (Argyris et al. 2008a), priming also reduced the expression of

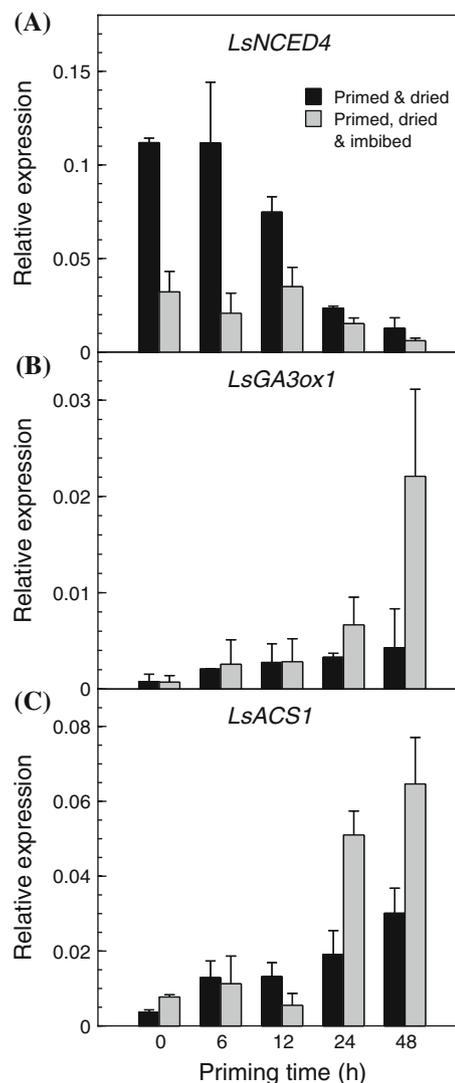


Fig. 7 Expression (mRNA abundance measured by quantitative RT-PCR) of *LsNCED4* (a), *LsGA3ox1* (b), and *LsACS1* (c) in Titan lettuce seeds that were primed for 0 (dry control seeds), 6, 12, 24 and 48 h. After osmopriming at 9°C in -1.25 MPa PEG 8000 for the indicated times, the seeds were rinsed, rapidly dried (2 h at 32°C), equilibrated at 9°C and 30% RH for 2 days, and then extracted (black bars). Control and primed seeds were subsequently imbibed in water at 35°C for 24 h, and then extracted for gene expression assays (grey bars). Note that the y-axes of the panels have different scales. Error bars represent standard error of 3 biological replicates

LsNCED4 in seeds of Salinas (Fig. 5a), NILs (Fig. 6a, b), and Titan (Fig. 7a), relative to the non-primed seeds of those genotypes imbibed at high temperatures. Thus, priming results in a reduced response of *LsNCED4* expression to imbibition at high temperature. As higher expression of *LsNCED4* is associated with increased ABA content and inhibition of germination (Argyris et al. 2008a), reduced expression of *LsNCED4* following priming likely results in lower ABA content and better germination at high temperatures relative to the non-primed seeds (Fig. 8).

The critical period for induction of thermoinhibition is within the first 8–12 h of imbibition at high temperature (Argyris et al. 2008a). Imbibition at a lower temperature during priming apparently allows germination processes to progress past this critical point, and subsequent exposure to high temperatures is not able to induce *LsNCED4* expression and prevent germination (Figs. 5a, 6a, 6b, 7a). This could be related to the developmental specificity of *LsNCED4* expression, which may be restricted to seed development and early imbibition. *LsNCED4* is the primary *NCED* family member expressed during late seed maturation in lettuce (H. Huo and K.J. Bradford, unpublished results), and the closest homolog of *LsNCED4* in Arabidopsis (*AtNCED6*) exhibits seed-specific expression (Lefebvre et al. 2006; Seo et al. 2006). *LsNCED4* mRNA abundance decreased during priming (Fig. 7a) as it does during imbibition in water at permissive temperatures (Argyris et al. 2008a). Thus, priming may allow the seeds to progress past the developmental point at which *LsNCED4* can be re-induced by high temperature (Fig. 8). Expression of *LsGA3ox1* and *LsACS1* exhibits the opposite

pattern, as they are not expressed in late seed maturation but are induced during imbibition or priming at low temperature (Fig. 7b, c; Argyris et al. 2008a). Imbibition at high temperature can initially repress expression of both genes (Fig. 6), but not after priming or after the escape time for induction of thermoinhibition (Fig. 8). This suggests that priming acts primarily by allowing the normal events of germination to advance while preventing embryo expansion due to the reduced water potential, rather than by inducing unique regulatory responses.

Reciprocal regulation of expression of ABA versus GA and ethylene biosynthetic genes in lettuce seed germination has been reported (Fig. 8; Gonai et al. 2004; Argyris et al. 2008a; Sawada et al. 2008a, b). The genetic evidence presented here suggests that regulation of *LsNCED4* expression and therefore of ABA levels may be a primary temperature-sensing mechanism, at least in these genetic backgrounds, which then triggers subsequent gene expression/repression and metabolic responses in other hormonal pathways. For example, ABA enhances the catabolism of GA and can inhibit the expression of GA biosynthetic genes in Arabidopsis and lettuce seeds (Gonai et al. 2004; Seo et al. 2006; Zentella et al. 2007; Sawada et al. 2008b; Toh et al. 2008). In addition, GA can reciprocally inhibit ABA biosynthesis and promote its catabolism (Oh et al. 2007; Sawada et al. 2008a). Gonai et al. (2004) reported that applications of fluridone (an ABA biosynthesis inhibitor) and GA together decreased the ABA content of lettuce seeds to a sufficiently low level to permit germination at 33°C in the dark. GA is required for lettuce seed germination at high temperatures even when ABA biosynthesis is inhibited (Endo et al. 2001; Argyris et al. 2008a). Reduced ethylene production is associated with the inhibition of germination of thermosensitive lettuce genotypes at high temperatures (Nascimento et al. 2000; Kozarewa et al. 2006; Matilla and Matilla-Vazquez 2008). Priming repressed expression of both Salinas and UC96US23 alleles of *LsNCED4* in NILs at 35°C, but expression of *LsGA3ox1* and *LsACS1* was still suppressed (either completely or partially) at this temperature (Fig. 6), suggesting that there may be direct effects of temperature on *LsGA3ox1* and *LsACS1* expression (Fig. 8). Previous results suggested that *LsGA3ox1* and *LsACS1* might be coordinately regulated (Argyris et al. 2008a); however, while the overall expression patterns are quite similar (Figs. 5, 7), their temperature sensitivities appear to be distinct (Fig. 6). Overall, regulation of the synthesis and action of ABA, GA and ethylene is apparently so tightly integrated that environmental influences result in coordinated reciprocal adjustments throughout the regulatory network (Fig. 8; Finch-Savage and Leubner-Metzger 2006; Oh et al. 2007; Finkelstein et al. 2008; Matilla and Matilla-Vazquez 2008).

Light is another major environmental factor influencing seed germination, and there is evidence that light and

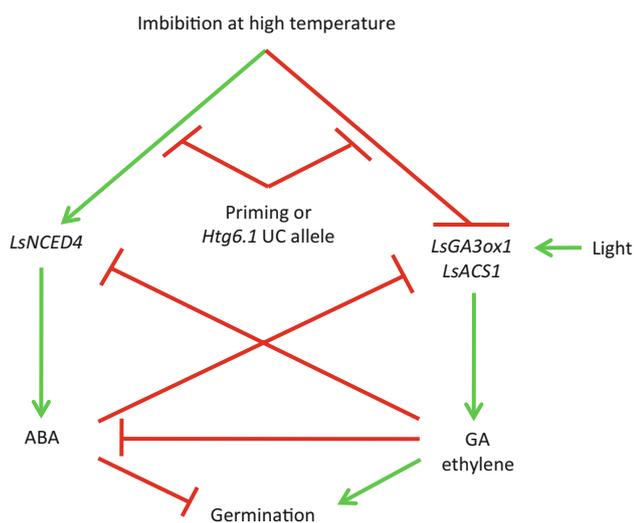


Fig. 8 Proposed model for regulation of thermoinhibition in lettuce seeds and its alleviation by priming. Arrows indicate promotion and bars inhibition of the process or gene expression

temperature interact through the phytochromes in regulating germination (Heschel et al. 2007). In lettuce seeds, the active phytochrome (Pfr) requirement for germination increases as the temperature increases (Fielding et al. 1992; Kristie and Fielding 1994) and the seeds become more sensitive to inhibition by ABA (Roth-Bejerano et al. 1999; Gonai et al. 2004). Pfr promotes GA biosynthesis (Toyomasu et al. 1998), and light altered both GA responsiveness and GA metabolism in Arabidopsis seeds (Oh et al. 2007). However, GA responsiveness was unaffected by red light treatment of lettuce seeds at 25°C, indicating that germination was mainly regulated by light via the control of endogenous levels of physiologically active GA (Sawada et al. 2008b). The priming treatments imposed here included exposure to fluorescent (red) light, which likely contributed to the strong response of *LsGA3ox1* expression to priming (Fig. 7). UC96US23 seeds require light for germination, and their thermotolerance is strictly dependent upon light, presumably to promote GA and/or ethylene biosynthesis (Argyris et al. 2008a). GA could substitute for light in promoting germination at high temperature (32°C) in the dark (Argyris et al. 2008a). Both progression past a developmental window for temperature-dependent *LsNCED4* expression and induction of *LsGA3ox1* and *LsACSI* expression by light during priming likely contribute to the priming effect (Fig. 8).

Conclusion

We have presented evidence that a genetic locus containing the ABA biosynthetic gene *LsNCED4* influences the responsiveness of lettuce seeds to priming with respect to the maximum temperature for germination. Introgression of this *Htg6.1* locus into a lettuce cultivar increased seed germination at high temperature and enhanced responsiveness to priming, making this treatment even more effective in alleviating thermoinhibition. Reduced expression of *LsNCED4* and increased expression of *LsGA3ox1* and *LsACSI* in primed seeds compared to non-primed seeds is associated with the increase in the maximum germination temperature. The expression of these ABA, GA, and ethylene biosynthetic genes could serve as molecular markers to optimize priming treatments. Developmental regulation of gene expression may contribute to the priming effect as the imbibition period allows seeds to progress past a germination checkpoint that is sensitive to thermoinhibition. One challenge for the future will be unraveling whether temperature acts only or primarily through effects on *LsNCED4* expression or whether multiple targets independently sense and respond to temperature. Regardless of the primary trigger(s), a complex network of changes in gene expression is associated with

germination responses to both priming and temperature, with the fate of germination or dormancy of individual seeds likely being determined by the relative balance between promotive and inhibitory signals.

Acknowledgments Richard Michelmore provided the RIL population, Allen Van Deynze and Richard Michelmore provided the high-density genetic map, Jason Argyris developed the NILs, Hamid Ashrafi and Maria Truco assisted with the QTL analyses, Peetambar Dahal helped with the quantitative RT-PCR assays, and Anthony Joudi assisted with the germination tests. This work was supported by the Western Regional Seed Physiology Research Group and the U.S. National Science Foundation (grant nos. 0421630 and 0820451) through the Compositae Genome Project (<http://compgenomics.ucdavis.edu>).

Open Access This article is distributed under the terms of the Creative Commons Attribution Noncommercial License which permits any noncommercial use, distribution, and reproduction in any medium, provided the original author(s) and source are credited.

References

- Argyris JM (2008) Natural variation and genetic analyses of seed and seedling traits in lettuce: discovery, confirmation and proposed role for a quantitative trait locus in regulating seed dormancy at high temperature. Ph.D. Thesis, University of California, Davis
- Argyris J, Truco MJ, Ochoa O, Knapp SJ, Still DW, Lenssen GM, Schut JW, Michelmore RW, Bradford KJ (2005) Quantitative trait loci associated with seed and seedling traits in *Lactuca*. *Theor Appl Genet* 111:1365–1376
- Argyris J, Dahal P, Hayashi E, Still DW, Bradford KJ (2008a) Genetic variation for lettuce seed thermoinhibition is associated with temperature-sensitive expression of abscisic acid, gibberellin and ethylene biosynthesis, metabolism and response genes. *Plant Physiol* 148:926–947
- Argyris J, Dahal P, Truco MJ, Ochoa O, Still DW, Michelmore RW, Bradford KJ (2008b) Genetic analysis of lettuce seed thermoinhibition. *Acta Hort* 782:23–33
- Benjamin LR (1990) Variation in time of seedling emergence within populations: a feature that determines individual growth and development. *Adv Agron* 44:1–25
- Berrie AMM (1966) Effect of temperature and light on germination of lettuce seeds. *Physiol Plant* 19:429–436
- Borthwick HA, Robbins WW (1928) Lettuce seed and its germination. *Hilgardia* 3:275–305
- Bradford KJ, Somasco OA (1994) Water relations of lettuce seed thermoinhibition: I. Priming and endosperm effects on base water potential. *Seed Sci Res* 4:1–10
- Bruggink GT (2004) Update on seed priming: from priming to pregermination, and back. *Seed Technol* 26:86–91
- Cantliffe DJ, Shuler KD, Guedes AC (1981) Overcoming seed thermodormancy in a heat-sensitive romaine lettuce by seed priming. *HortScience* 16:196–198
- Cantliffe DJ, Fischer JM, Nell TA (1984) Mechanism of seed priming in circumventing thermodormancy in lettuce. *Plant Physiol* 75:290–294
- Cantliffe DJ, Sung Y, Nascimento WM (2000) Lettuce seed germination. *Hortic Rev* 224:229–275
- Chen F, Bradford KJ (2000) Expression of an expansin is associated with endosperm weakening during tomato seed germination. *Plant Physiol* 124:1265–1274
- Contreras S, Bennett MA, Metzger JD, Tay D, Nerson H (2009) Red to rar-red ratio during seed development affects lettuce seed germinability and longevity. *HortScience* 44:130–134

- Cooley MB, Yang H, Dahal P, Mella RA, Downie AB, Haigh AM, Bradford KJ (1999) Vacuolar H⁺-ATPase is expressed in response to gibberellin during tomato seed germination. *Plant Physiol* 121:1339–1347
- de Castro RD, van Lammeren AAM, Groot SPC, Bino RJ, Hilhorst HWM (2000) Cell division and subsequent radicle protrusion in tomato seeds are inhibited by osmotic stress but DNA synthesis and formation of microtubular cytoskeleton are not. *Plant Physiol* 122:327–335
- Dutta S, Bradford KJ (1994) Water relations of lettuce seed thermoinhibition. II. Ethylene and endosperm effects on base water potential. *Seed Sci Res* 4:11–18
- Dutta S, Bradford KJ, Nevins DJ (1997) Endo-beta-mannanase activity present in cell wall extracts of lettuce endosperm prior to radicle emergence. *Plant Physiol* 113:155–161
- Endo K, Yoshioka T, Gonai T, Satoh S, Hashiba T (2001) Induction of seed germination at high temperatures by herbicides which inhibit carotenoid biosynthesis. *Tohoku Weed J* 1:3–6
- Fielding A, Kristie DN, Dearman P (1992) The temperature of Pfr action governs the upper temperature limit for germination in lettuce. *Photochem Photobiol* 56:623–627
- Finch-Savage WE, Leubner-Metzger G (2006) Seed dormancy and the control of germination. *New Phytol* 171:501–523
- Finkelstein R, Reeves W, Ariizumi T, Steber C (2008) Molecular aspects of seed dormancy. *Ann Rev Plant Biol* 59:387–415
- Gallardo K, Job C, Groot SPC, Puype M, Demol H, Vandekerckhove J, Job D (2001) Proteomic analysis of *Arabidopsis* seed germination and priming. *Plant Physiol* 126:835–848
- Gao YP, Young L, Bonham-Smith P, Gusta LV (1999) Characterization and expression of plasma and tonoplast membrane aquaporins in primed seed of *Brassica napus* during germination under stress conditions. *Plant Mol Biol* 40:635–644
- Gonai T, Kawahara S, Tougou M, Satoh S, Hashiba T, Hirai N, Kawaide H, Kamiya Y, Yoshioka T (2004) Abscisic acid in the thermoinhibition of lettuce seed germination and enhancement of its catabolism by gibberellin. *J Exp Bot* 55:111–118
- Gray D (1975) Effects of temperature on the germination and emergence of lettuce (*Lactuca sativa* L.) varieties. *Hortic Sci* 50:349–361
- Halmer P, Bewley JD, Thorpe TA (1975) Enzyme to break down lettuce endosperm cell wall during gibberellin-induced and light-induced germination. *Nature* 258:716–718
- Heschel MS, Selby J, Butler C, Whitlam GC, Sharrock RA, Donohue K (2007) A new role for phytochromes in temperature-dependent germination. *New Phytol* 174:735–741
- Heydecker W, Coolbear P (1977) Seed treatments for improving performance—survey and attempted prognosis. *Seed Sci Technol* 5:353–425
- Heydecker W, Higgins J, Gulliver RL (1973) Accelerated germination by osmotic seed treatment. *Nature* 246:42–44
- Job C, Kersulec A, Ravaio L, Chareyre S, Pepin R, Job D (1997) The solubilization of the basic subunit of sugarbeet seed 11-S globulin during priming and early germination. *Seed Sci Res* 7:225–243
- Kozarewa I, Cantliffe DJ, Nagata RT, Stoffella PJ (2006) High maturation temperature of lettuce seeds during development increased ethylene production and germination at elevated temperatures. *J Am Soc Hortic Sci* 131:564–570
- Kristie DN, Fielding A (1994) Influence of temperature on the Pfr level required for germination in lettuce cv. Grand Rapids. *Seed Sci Res* 4:19–25
- Kucera B, Cohn MA, Leubner-Metzger G (2005) Plant hormone interactions during seed dormancy release and germination. *Seed Sci Res* 15:281–307
- Lefebvre V, North H, Frey A, Sotta B, Seo M, Okamoto M, Nambara E, Marion-Poll A (2006) Functional analysis of *Arabidopsis NCED6* and *NCED9* genes indicates that ABA synthesised in the endosperm is involved in the induction of seed dormancy. *Plant J* 45:309–319
- Ligterink W, Kodde J, Lammers M, de Maagd R, Hilhorst H (2007) Molecular dissection of seed priming. Translational seed biology: from model systems to crop improvement, Davis, CA. <http://www.plantsciences.ucdavis.edu/Symposium/2007/>
- Matilla AJ, Matilla-Vazquez MA (2008) Involvement of ethylene in seed physiology. *Plant Sci* 175:87–97
- McDonald MB (2000) Seed priming. In: Black M, Bewley JD (eds) *Seed technology and its biological basis*. Sheffield Academic Press, Sheffield, pp 287–325
- Müller K, Tintelnot S, Leubner-Metzger G (2006) Endosperm-limited Brassicaceae seed germination: abscisic acid inhibits embryo-induced endosperm weakening of *Lepidium sativum* (cress) and endosperm rupture of cress and *Arabidopsis thaliana*. *Plant Cell Physiol* 47:864–877
- Nascimento WM, Cantliffe DJ, Huber DJ (2000) Thermotolerance in lettuce seeds: association with ethylene and endo-beta-mannanase. *J Am Soc Hortic Sci* 125:518–524
- Nascimento WM, Cantliffe DJ, Huber DJ (2001) Endo-beta-mannanase activity and seed germination of thermosensitive and thermotolerant lettuce genotypes in response to seed priming. *Seed Sci Res* 11:255–264
- Nonogaki H, Morohashi Y (1999) Temporal and spatial pattern of the development of endo-beta-mannanase activity in germinating and germinated lettuce seeds. *J Exp Bot* 50:1307–1313
- Nonogaki H, Chen F, Bradford KJ (2007) Mechanisms and genes involved in germination *sensu stricto*. In: Bradford KJ, Nonogaki H (eds) *Seed development, dormancy and germination*. Blackwell Publishing, Oxford, pp 264–304
- Oh E, Yamaguchi S, Hu JH, Yusuke J, Jung B, Paik I, Lee HS, Sun TP, Kamiya Y, Choi G (2007) PIL5, a phytochrome-interacting bHLH protein, regulates gibberellin responsiveness by binding directly to the GAI and RGA promoters in *Arabidopsis* seeds. *Plant Cell* 19:1192–1208
- Reynolds T, Thompson PA (1973) Effects of kinetin, gibberellins and (±) abscisic acid on the germination of lettuce (*Lactuca sativa*). *Physiol Plant* 28:516–522
- Roth-Bejerano N, Sedee NJA, van der Meulen RM, Wang M (1999) The role of abscisic acid in germination of light-sensitive and light-insensitive lettuce seeds. *Seed Sci Res* 9:129–134
- Saini H, Consolacion E, Bassi P, Spencer M (1986) Requirement for ethylene synthesis and action during relief of thermoinhibition of lettuce seed germination by combinations of gibberellin acid, kinetin, and carbon dioxide. *Plant Physiol* 81:950–953
- Saini HS, Consolacion ED, Bassi PK, Spencer MS (1989) Control processes in the induction and relief of thermoinhibition of lettuce seed germination: actions of phytochrome and endogenous ethylene. *Plant Physiol* 90:311–315
- Sawada Y, Aoki M, Nakaminami K, Mitsuhashi W, Tatematsu K, Kushihiro T, Koshihara T, Kamiya Y, Inoue Y, Nambara E, Toyomasu T (2008a) Phytochrome- and gibberellin-mediated regulation of abscisic acid metabolism during germination of photoblastic lettuce seeds. *Plant Physiol* 146:1386–1396
- Sawada Y, Katsumata T, Kitamura J, Kawaide H, Nakajima M, Asami T, Nakaminami K, Kurahashi T, Mitsuhashi W, Inoue Y, Toyomasu T (2008b) Germination of photoblastic lettuce seeds is regulated via the control of endogenous physiologically active gibberellin content, rather than of gibberellin responsiveness. *J Exp Bot* 59:3383–3393
- Schopfer P, Plachy C (1984) Control of seed germination by abscisic acid. 2. Effect on embryo water uptake in *Brassica napus* L. *Plant Physiol* 76:155–160
- Seo M, Hanada A, Kuwahara A, Endo A, Okamoto M, Yamauchi Y, North H, Marion-Poll A, Sun T-p, Koshihara T, Kamiya Y,

- Yamaguchi S, Nambara E (2006) Regulation of hormone metabolism in Arabidopsis seeds: phytochrome regulation of abscisic acid metabolism and abscisic acid regulation of gibberellin metabolism. *Plant J* 48:354–366
- Soeda Y, Konings M, Vorst O, van Houwelingen A, Stoop GM, Maliepaard CA, Kodde J, Bino RJ, Groot SPC, van der Geest AHM (2005) Gene expression programs during *Brassica oleracea* seed maturation, osmopriming, and germination are indicators of progression of the germination process and the stress tolerance level. *Plant Physiol* 137:354–368
- Speer HL (1974) Some aspects of function of endosperm during germination of lettuce seeds. *Can J Bot* 52:1117–1121
- Stam P (1995) JoinMap 2.0 deals with all types of plant mapping populations. *Plant genome III abstracts*. <http://www.intl-pag.org/3/abstracts/47pg3.html>
- Sung Y, Cantliffe DJ, Nagata R (1998) Using a puncture test to identify the role of seed coverings on thermotolerant lettuce seed germination. *J Am Soc Hortic Sci* 123:1102–1106
- Sung Y, Cantliffe DJ, Nagata RT, Nascimento WM (2008) Structural changes in lettuce seed during germination at high temperature altered by genotype, seed maturation temperature, and seed priming. *J Am Soc Hortic Sci* 133:300–311
- Takeba G, Matsubara S (1979) Measurement of the growth potential of the embryo in New York lettuce seed under various combinations of temperature, red light and hormones. *Plant Cell Physiol* 20:51–61
- Tan BC, Joseph LM, Deng WT, Liu LJ, Li QB, Cline K, McCarty DR (2003) Molecular characterization of the *Arabidopsis* 9-cis epoxy-carotenoid dioxygenase gene family. *Plant J* 35:44–56
- Tarquis AM, Bradford KJ (1992) Prehydration and priming treatments that advance germination also increase the rate of deterioration of lettuce seeds. *J Exp Bot* 43:307–317
- Toh S, Imamura A, Watanabe A, Nakabayashi K, Okamoto M, Jikumaru Y, Hanada A, Aso Y, Ishiyama K, Tamura N, Iuchi S, Kobayashi M, Yamaguchi S, Kamiya Y, Nambara E, Kawakami N (2008) High temperature-induced abscisic acid biosynthesis and its role in the inhibition of gibberellin action in Arabidopsis seeds. *Plant Physiol* 146:1368–1385
- Toorop PE, van Aelst AC, Hilhorst HWM (2000) The second step of the biphasic endosperm cap weakening that mediates tomato (*Lycopersicon esculentum*) seed germination is under control of ABA. *J Exp Bot* 51:1371–1379
- Toyomasu T, Kawaide H, Mitsuhashi W, Inoue Y, Kamiya Y (1998) Phytochrome regulates gibberellin biosynthesis during germination of photoblastic lettuce seeds. *Plant Physiol* 118:1517–1523
- Truco MJ, Antonise R, Lavelle D, Ochoa O, Kozik A, Witsenboer H, Fort SB, Jeuken MJW, Kesseli RV, Lindhout P, Michelmore RW, Peleman J (2007) A high-density, integrated genetic linkage map of lettuce (*Lactuca* spp.). *Theor Appl Genet* 115:735–746
- Valdes VM, Bradford KJ, Mayberry KS (1985) Alleviation of thermodormancy in coated lettuce seeds by seed priming. *HortScience* 20:1112–1114
- van Leeuwen H, Caldwell DG, Stoffel K, Chen F, Kozik A, Truco MJ, Michelmore RW, Van Deynze AE (2007) GeneChip for massively parallel genotyping and gene expression analysis. *Plant and Animal Genome XV*, San Diego
- van Leeuwen H, Stoffel K, Kozik A, Cui X, Ashrafi H, McHale L, Lavelle D, Wong G, Chen F, Truco MJ, Van Deynze A, Michelmore RW (2009) High-density mapping of the lettuce genome with SFP markers in over 15,000 unigenes. In *Plant and Animal Genome XVII*, San Diego, CA
- Weges R (1987) Physiological analysis of methods to relieve dormancy of lettuce seeds. Ph.D. Thesis, Agricultural University, Wageningen, The Netherlands
- Wu CT, Leubner-Metzger G, Meins F, Bradford KJ (2001) Class I beta-1,3-glucanase and chitinase are expressed in the micropylar endosperm of tomato seeds prior to radicle emergence. *Plant Physiol* 126:1299–1313
- Zentella R, Zhang ZL, Park M, Thomas SG, Endo A, Murase K, Fleet CM, Jikumaru Y, Nambara E, Kamiya Y, Sun TP (2007) Global analysis of DELLA direct targets in early gibberellin signaling in Arabidopsis. *Plant Cell* 19:3037–3057

Supplemental data

Table S1. Statistical analysis of the overall priming effect on seeds of the RIL population.

The germination percentage values at each temperature between 27 and 35°C were transformed by the probit method to normalize variances, and statistical analysis was conducted using the General Linear Model of the SAS statistical program (SAS Institute Inc., Cary, NC, USA).

Source	DF	Type III SS	MS	F value	Pr > F
Replicate	1	0.54	0.54	2.50	0.1141 NS
RIL	88	5933.54	67.43	310.08	<0.0001
Treatment	1	235.14	235.14	1081.37	<0.0001
Temperature	8	3450.54	431.32	1983.55	<0.0001
RIL x Treatment	88	215.68	2.45	11.27	<0.0001
RIL x Temperature	704	1303.47	1.85	8.51	<0.0001
Treat x Temp	8	27.50	3.44	15.81	<0.0001
RIL x Treat x Temp	704	333.56	0.47	2.18	<0.0001

DF = Degrees of freedom

SS = Sum of squares

MS = Mean square

NS = non-significant

Table S2. Covariate analysis of the priming effect on germination of seeds of the RIL population at temperatures between 27 and 35°C.

Source	DF	Type III SS	MS	F value	Pr > F
X (covariant, allele effect)	1	156.04	156.04	517.38	<0.0001
Replicate	1	0.52	0.52	1.72	0.2103 NS
RIL	88	4455.67	50.63	272.38	<0.0001
Treatment	1	201.28	201.28	812.33	<0.0001
Temperature	8	2779.89	347.50	1583.62	<0.0001
RIL x Treatment	88	198.68	2.26	10.17	<0.0001
RIL x Temperature	704	1303.47	1.85	8.51	<0.0001
Treat x Temp	8	22.99	2.87	12.31	<0.0001
RIL x Treat x Temp	704	303.22	0.43	4.11	<0.0001

DF = Degrees of freedom

SS = Sum of squares

MS = Mean square

NS = Non-Significant

Table S3. Statistical analysis of the overall priming effect on seeds of NILs 2, 27, and 86.

Each NIL was homozygous for either the Salinas or the UC96US23 alleles in a genomic region containing *Htg6.1*. The germination percentage values of the control and the primed seeds were transformed by the probit method to normalize variances, and statistical analysis was conducted using the General Linear Model of the SAS statistical program (SAS Institute Inc., Cary, NC, USA).

Source	DF	Type III SS	MS	F value	Pr > F
Temperature	8	1788.90	223.61	615.33	<0.0001
Treatment	1	308.18	308.18	848.03	<0.0001
NIL	2	23.90	11.95	32.89	<0.0001
Allele	1	520.55	520.55	1432.43	<0.0001
Temp x Treatment	8	102.11	12.76	35.12	<0.0001
Temp x NIL	16	8.56	0.53	1.47	0.1027 NS
Temp x Allele	8	80.48	10.06	27.68	<0.0001
Treatment x NIL	2	5.43	2.71	7.47	0.0006
Treatment x Allele	1	64.76	64.76	178.2	<0.0001
NIL x Allele	2	2.18	1.09	2.99	0.505 NS
Temp x Treatment x NIL	16	10.76	0.67	1.85	0.0216
Temp x Treatment x Allele	8	24.44	3.05	8.41	<0.0001
Treatment x NIL x Allele	2	2.24	1.12	3.08	0.0464
Temp x Treat x NIL x Allele	32	17.10	0.53	1.47	0.0455

DF = Degrees of freedom

SS = Sum of squares

MS = Mean square

NS = Non-significant